

Human Umbilical Vein Endothelial

Cells

Product sheet, catalog n° CTICC1.14

General Information

- Organism: Human (Homo sapiens)
- Tissue: Umbilical cord
- Cell Type: Umbilical vein endothelial cells from single donor
- Location: Stem cell extracted from the vein of umbilical cord
- Gender: Male or Female (see Certificate of Analysis)
- Age: Newborn (see Certificate of Analysis)
- Phototype (Fitzpatrick scale): (see Certificate of Analysis)

Cell Characteristics

- Cell properties: Adherent
- Morphology: Cobblestone shape
- Isolation: Enzymatic dissociation
- Cell passage: P0 to P2 (see Certificate of Analysis)
- Minimum number of population doublings: 15
- Cell viability: Minimum 80% viability when thawed from cryopreservation
- Cell conditioning: Supplied as vials of 1.10⁶ cells
- Cryopreservation medium: Frozen with 90% serum-free cryopreservation medium + 10% DMSO
- Storage condition: Liquid nitrogen
- Batch specific information: Included in the Certificate of Analysis

Safety and Quality Control

- Biosafety level: 1
- **Contamination:** Use mandatory laboratory protections and handle with care tissues and cells derived from human samples to avoid any contamination of the operator.
- Viral testing: Negative for HIV, HBV, HCV
- Sterility testing: Negative for mycoplasma, bacteria and yeasts

Handling upon delivery and storage

- Check that the containers are intact and free of damage
- If cells are not used immediately, place the vials at -150°C or below upon delivery

Thawing and culturing procedure for frozen cells

- 1 Add 0,12 ml per cm² of medium to the culture vessel
- 2 Add 13 ml of PBS solution to a 15 ml conical tube, and warm in a water bath to 37 °C
- 3 Thaw cryovial by swirling in a water bath at 37 °C. As soon as the content has thawed, start step 4
- 4 Once thawed, add the cell suspension to the warmed PBS in sterile conditions
- 5 Spin the tube at 250 g for 7 minutes to pellet the cells
- 6 Resuspend the cells in the appropriate volume of recommended medium





- 7 Seed the cells in the culture vessel at a concentration of 5 000 to 10 000 cells per cm²
- 8 Incubate at 37°C, 5% CO2 atmosphere, 95% humidity
- 9 After 24 hours of incubation, change the medium to remove any debris
- 10 Continue to incubate and change the medium every 3-4 days.

Subculturing

- 1- Start subculturing when cells reached 70%-80% confluence
- 2 Preheat TrypLE (non toxic for cell trypsin substitute) and recommended medium.
- 3 Remove the medium from the flask
- 4 Wash the cells quickly with PBS without Ca2+ Mg2+
- 5 Add 0,06 mL per cm² of TrypLE for 5-15 min at 37 °C in the incubator
- 6 Remove the cells from the flask by pipetting several times and wash the flask with recommended medium for remaining cells
- 7 Centrifuge the cells in recommended medium at 250 g for 7 minutes to pellet the cells
- 8 Remove the supernatant and resuspend the pellet in recommended medium
- 9 Seed the cells in the culture vessel at a concentration of 5 000 cells per cm²
- 10 Incubate at 37 °C, 5% CO₂ atmosphere, 95% humidity
- 11 After 24 hours of incubation, change half of the medium to remove any debris
- 12 Continue to incubate and change half of the medium every 3-4 days.

Associated products

- CTICC.1.11: Human Amniotic Membrane Epithelial Stem Cells, Cryopreserved, 10⁶ cells
- CTIINT.1.5: Human Amniotic Membrane, Fresh, 1cm²
- CTIINT.1.1: Human Placenta, Full Thickness, Fresh, 1cm³
- CTIINT.1.9: Human Umbilical Cord, Fresh, 1cm

Provisions

- Cells and tissues are intended for **research use only** and shall not be used for human trials, animal trials, or diagnostics.
- **Consent:** the original tissues have been obtained after informed consent of the patient under the provisions required by French Law.
- Primary Human cells are not immortalised cell lines and may not be continually subcultured.