

Juvenile Human Melanocytes



Product sheet, catalog n° CTICC.1.3.3

General Information

• Organism: Human (Homo sapiens)

Tissue: Skin

Cell Type: Skin Melanocytes from single donor

• Location: Foreskin or other part (see Certificate of Analysis)

Gender: Male or Female (see Certificate of Analysis)

Age: 1 – 17 years (see Certificate of Analysis)

Phototype: (see Certificate of Analysis)

Cell Characteristics

- Cell properties: Adherent
- Morphology: Stellar or dendritic shaped
- Isolation: Enzymatic dissociation
- Cell passage: P1 to P4 (see Certificate of Analysis)
- Minimum number of population doublings: 15
- Cell viability: Minimum 70% viability when thawed from cryopreservation
- Cell conditioning: Supplied as vials of 1M cells
- Cryopreservation medium: Frozen with 90% serum-free cryopreservation medium + 10% DMSO
- Storage condition: Liquid nitrogen
- Batch specific information: (see Certificate of Analysis)

Safety and Quality Control

- Biosafety level: 1
- Contamination: Use mandatory laboratory protections and handle with care tissues and cells derived from human samples to avoid any contamination of the operator
- Viral testing: negative for HIV, HBV, HCV
- Sterility testing: Negative for mycoplasma, bacteria and yeasts

Handling upon delivery and storage

- · Check that the containers are intact and free of damage
- If cells are not used immediately, place the vials at -150°C or below upon delivery

Growth medium

Recommended medium reference: CTIGM.Melano: Growth Medium for Melanocytes



Thawing and culturing procedure for frozen cells

- 1 Add 0.12 ml per cm² of medium to the culture vessel (Recommended: 9 mL in T75 flask)
- 2 Add 13 ml of PBS solution to a 15 ml conical tube, and warm in a water bath to 37 °C
- 3 Thaw cryovial by swirling in a water bath at 37 °C. As soon as the content has thawed, start step 4
- 4 Once thawed, add the cell suspension to the warmed PBS in sterile conditions
- 5 Spin the tube at 250 g for 7 minutes to pellet the cells
- 6 Resuspend the cells in the appropriate volume of recommended medium
- 7 Seed the cells in the culture vessel at a concentration of 5 000 to 10 000 cells per cm²
- 7 Incubate at 37°C, 5% CO2 atmosphere, 95% humidity
- 8 After 24 hours of incubation, change the medium to remove any debris
- 9 Continue to incubate and change the medium every 3-4 days

Subculturing

- 1- Start subculturing when cells reached 80%-90% confluence
- 2 Preheat TrypLE (non-toxic for cell trypsin substitute) and recommended medium
- 3 Remove the medium from the flask
- 4 Wash the cells quickly with PBS without Ca2+ Mg2+
- 5 Add 0,06mL per cm² of TrypLE for 3-4 min at 37 °C in the incubator
- 6 Remove the cells from the flask by pipetting several times and wash the flask with recommended medium for remaining cells
- 7 Centrifuge the cells in recommended medium at 250 g for 7 minutes to pellet the cells
- 8 Remove the supernatant and resuspend the pellet in recommended medium
- 9 Seed the cells in the culture vessel at a concentration of 5 000 cells per cm²
- 10 Incubate at 37 °C, 5% CO2 atmosphere, 95% humidity
- 11 After 24 hours of incubation, change half of the medium to remove any debris
- 12 Continue to incubate and change the medium every 3-4 days

Associated products

- CTICC.1.1.3: Juvenile Human Dermal Fibroblasts, Cryopreserved, 10⁶ cells
- CTICC.1.2.3: Juvenile Human Keratinocytes, Cryopreserved, 10⁶ cells
- SKIN BIOPSIES: Fresh, Flash Frozen, FFPE, OCT-embedded

Provisions

- Cells and tissues are intended for **research use only** and shall not be used for human trials, animal trials, or diagnostics.
- Consent: the original tissues have been obtained after informed consent of the patient under the provisions required by French Law.
- Primary Human cells are not immortalised cell lines and may not be continually subcultured.