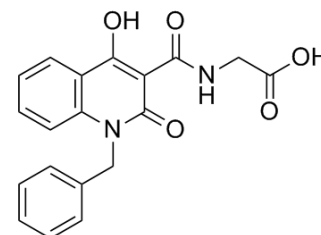


IOX2

Cat. No.:	HY-15468		
CAS No.:	931398-72-0		
Molecular Formula:	C ₁₉ H ₁₆ N ₂ O ₅		
Molecular Weight:	352.34		
Target:	HIF/HIF Prolyl-Hydroxylase		
Pathway:	Metabolic Enzyme/Protease		
Storage:	Powder	-20°C	3 years
		4°C	2 years
	In solvent	-80°C	6 months
		-20°C	1 month



SOLVENT & SOLUBILITY

In Vitro	DMSO : 25 mg/mL (70.95 mM; Need ultrasonic)				
		Solvent Concentration	Mass 1 mg	5 mg	10 mg
	Preparing Stock Solutions	1 mM	2.8382 mL	14.1908 mL	28.3817 mL
		5 mM	0.5676 mL	2.8382 mL	5.6763 mL
10 mM		0.2838 mL	1.4191 mL	2.8382 mL	
Please refer to the solubility information to select the appropriate solvent.					
In Vivo	1. Add each solvent one by one: 10% DMSO >> 40% PEG300 >> 5% Tween-80 >> 45% saline Solubility: ≥ 2.5 mg/mL (7.10 mM); Clear solution				

BIOLOGICAL ACTIVITY

Description	IOX2 is a specific prolyl hydroxylase-2 (PHD2) inhibitor with IC ₅₀ of 22 nM.
IC₅₀ & Target	IC ₅₀ : 22 nM (PHD2) ^[1]
In Vitro	IOX2 is at least 2-5000 fold selective, as judged by IC ₅₀ values, for PHD2 over the KDMs and Factor Inhibiting HIF(FIH) ^[1] . IOX2 significantly upregulates the transcription of VEGF-A and BNIP3 in normal human epidermal keratinocytes (NHEK) and normal human dermal fibroblasts (NHDF) when grown under normoxia and hypoxia. IOX2 efficiently promotes HIF-1α stability, nuclear translocation, and target gene expression in keratinocytes and fibroblasts. In addition, IOX2 significantly upregulates biosynthesis and transcription of VEGF-A and BNIP3 in sulfur mustard (SM)-exposed NHEK and NHDF grown under hypoxia. These results suggest that application of IOX2 is useful for restoring of SM-affected HIF-1α stability and signaling activity in keratinocytes and fibroblasts ^[2] . MCE has not independently confirmed the accuracy of these methods. They are for reference only.

In Vivo	<p>To investigate the utility of IOX2 as in vivo functional probes, IOX2 is tested to upregulate HIF signaling in a whole organism, that is, transgenic zebrafish (<i>Danio rerio</i>). Because the expression of the PHD3 encoding gene is regulated by HIF in humans and zebrafish, PHD3 levels are a readout of HIF activity. A zebrafish hypoxia reporter line is generated expressing GFP with the <i>phd3</i> promoter elements. Transgenic wild-type embryos at 3 days postfertilization treated with compounds (10 μM) for 2 days displayed clear increase in <i>phd3:EGFP</i> expression in the liver, relative to controls. Significant increases in GFP levels are observed with IOX2^[1].</p> <p>MCE has not independently confirmed the accuracy of these methods. They are for reference only.</p>
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PROTOCOL

Kinase Assay ^[1]	<p>Inhibition assays are carried out in 384-well white ProxiPlates in 10 μL of reaction volume. Standard reaction mixtures consisted of the compound (in 2% DMSO final concentration), enzyme mix (0.001 μM of PHD2, 10 μM of Fe(II), 100 μM of ascorbate) and peptide mix (0.06 μM of biotinylated C-terminal oxygen dependent degradation domain (CODD) peptide, 2 μM of 2OG) in 50 mM HEPES pH 7.5, 0.01% Tween-20 and 0.1% BSA buffer. Compounds (e.g., IOX2) are preincubated with the enzyme mix for 15 min before being incubated with peptide mix for 10 min at 22°C. Each reaction is quenched with 5 μL of 30 mM EDTA. The bead mix containing AlphaScreen beads is preincubated for 1h with a rabbit monoclonal antibody selective for hydroxy-HIF1α (Pro564) and are added to the wells for a further 1 h at 22°C. The plates are then analyzed with an Envision plate reader. The IC₅₀ values are calculated using nonlinear regression with normalized dose-response fit using Prism GraphPad (n\geq3)^[1].</p> <p>MCE has not independently confirmed the accuracy of these methods. They are for reference only.</p>
Cell Assay ^[1]	<p>Both VHL-defective (renal carcinoma cells with an empty vector, RCC4) and VHL-competent cells human embryonic kidney HEK293T, osteosarcoma U2OS and RCC4/VHLHA (RCC4 stably transfected with C-terminal HA-tagged wt VHL) are used. Cells are treated with DMSO (control) and tested compounds (e.g., IOX2) (dissolved in DMSO except for DMOG which is dissolved in PBS and added directly to culture medium) for 4-5 h. Cell extracts are probed with antibodies to hydroxy-Pro564 (CODD-OH) and hydroxy-Asn803 (CAD-OH). HIF-1α band intensities are used to normalize hydroxylation signals^[1].</p> <p>MCE has not independently confirmed the accuracy of these methods. They are for reference only.</p>
Animal Administration ^[1]	<p>Zebrafish^[1]</p> <p>Phd3:gfp^{sh144/sh144} fish (<i>Danio rerio</i>) are incrossed to produce <i>phd3:gfp^{sh144/sh144}</i> embryos, these are raised at 28°C in E3 medium. The <i>phd3:gfp^{sh144/sh144}</i> line is a hypoxia reporter line created by BAC recombination of the <i>phd3</i> reporter GFP construct. At 3 days post fertilization, potential inhibitors (e.g., IOX2) are added in fresh medium at 10 μM in 1% DMSO. The embryos are incubated with the compounds for a further 48 h. Embryos are anesthetized at 5 dpf by immersion in tricaine. Lateral view images of the embryos are taken using a fluorescent dissecting stereomicroscope (both bright-field and fluorescent). Fluorescent images are analyzed using Image J.</p> <p>MCE has not independently confirmed the accuracy of these methods. They are for reference only.</p>

CUSTOMER VALIDATION

- Oxid Med Cell Longev. 2020 Jan 4;2020:4909103.
- Patent. US20180263995A1.
- Hypoxia (Auckl). 2016 Nov 8;4:147-159.
- Harvard Medical School LINCS LIBRARY

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REFERENCES

[1]. Chowdhury R, et al. Selective small molecule probes for the hypoxia inducible factor (HIF) prolyl hydroxylases. ACS Chem Biol. 2013 Jul 19;8(7):1488-96.

[2]. Deppe J, et al. Impairment of hypoxia-induced HIF-1 α signaling in keratinocytes and fibroblasts by sulfur mustard is counteracted by a selective PHD-2 inhibitor. Arch Toxicol. 2016 May;90(5):1141-50.

Caution: Product has not been fully validated for medical applications. For research use only.

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