

Streptomycin ELISA Test Kit

Catalogue Number. LT2A42001AYSL

Principle

This test kit is based on the competitive enzyme immunoassay for the detection of Streptomycin in the sample. The coupling antigen is pre-coated on the micro well stripes. The Streptomycin in the sample and the coupling antigens pre-coated on the micro well stripes compete for anti Streptomycin antibodies. After the addition of the enzyme conjugate, the TMB substrate is added for coloration. The optical density (OD) value of the testing sample has a negative correlation with the Streptomycin concentration in the sample. This value is compared to the standard curve and the Streptomycin concentration is subsequently obtained.

Technical specifications

Sensitivity: 0.1 ppb

Incubation Temperature: 25°C

Incubation Time: 30min—30min—15min

Detection limit: Chicken 1 ppb, Chicken liver, milk 4 ppb, Honey, Royal jelly 2 ppb

Recovery rate: Milk 85±22%, Chicken 80±17%, Honey, Royal jelly 75±19%

Cross-reaction rate: Streptomycin 100%, Dihydrostreptomycin 108%, Kalamycin<0.1%, Gentamycin<0.1%

Components

1	Micro-well strips	12 strips with 8 removable wells each	
2	Standard solution (1mL each)	0ppb, 0.1ppb, 0.3ppb, 0.9ppb, 2.7ppb, 8.1ppb	
3	High-concentration standard, 1ml	1ppm	red cap
4	Enzyme conjugate	11ml	red cap
5	Antibody working solution	5.5ml	blue cap
6	Substrate A	6ml	white cap
7	Substrate B	6ml	black cap
8	Stop solution	6ml	yellow cap
9	20× concentrated washing buffer	40 ml	white cap
10	5× concentrated redissolving solution	50ml	yellow cap

Materials required but not provided

Equipments: microplate reader, printer, homogeniser, nitrogen-drying device, vortex, centrifuge, measuring pipets, balance (a reciprocal sensibility of 0.01 g).

Micropipettors: single-channel 20~200 µL, 100~1000 µL; and multi-channel 300 µL;

Reagents: NaOH, Na₂HPO₄·12H₂O, NaH₂PO₄·2H₂O, N-hexane, Dichloromethane(CH₂Cl₂), Acetonitrile(CH₃CN), Phosphoric acid(H₃PO₄).

Sample pre-treatment

Instructions

- 1) Only the disposable tips can be used for the experiments and the tips must be changed when used for different reagents;
- 2) Before the experiment, each experimental equipment must be clean and should be re- cleaned if necessary, in order to avoid the contamination that interferes with the experimental results.

Solution preparation before sample pre-treatment

- 1) 0.05M PB Buffer: weigh 12.9g Na₂HPO₄· 12H₂O and 2.175g NaH₂PO₄· 2H₂O, add 1000mL deionized water to dissolve.
- 2) 0.04M Phosphoric acid(H₃PO₄) (for honey preparation): take 1mL concentrated Phosphoric acid(H₃PO₄), add deionized water to 360mL.
- 3) 1M NaOH solution (for honey preparation): weigh 4g NaOH, add deionized water to 100mL.
- 4) Redissolving solution: the 5×concentrated redissolving solution is diluted with deionized water at 1:4 (1mL concentrated redissolving solution + 4mL deionized water), the diluted Redissolving solution can store at 4°C for one month.

Samples preparation

Tissue (chicken, pork, beef, lamb etc.)

- 1 Take 2±0.05 g homogenized sample(remove fat), add 8mL 0.05M PB Buffer, shake for 5min, put it at 56°C water bath for 30min;
- 2 Centrifuge at above 4000 r/min at room temperature for 10 min.
- 3 Take 1ml up-layer clear liquid into a new vessel, add 1mL N-hexane, mix it evenly, centrifuge at above 4000 r/min at room temperature for 5min.
- 4 Remove up-layer organic phase, take 50ul down-layer liquid, add 450µL the diluted redissolving solution, mix properly for 30 seconds.
- 5 Take 50 µL for analysis.

Fold of dilution of sample: 40 Detection limit: 4ppb

Honey, Royal jelly

- 1 Weigh 2±0.05g honey sample, add 4mL 0.04M Phosphoric acid(H₃PO₄), shake until dissolved fully, centrifuge at above 4000 r/min at room temperature (20-25 °C) for 5min, until clear(honey sample do not need centrifuge, it can direct to step 2).
- 2 Add 450ul 1M NaOH and adjust the PH value to between 7 and 9 (Royal jelly need to transfer all up-layer clear liquid into a new clean centrifuge tube, and adjust the PH value to between 7 and 9), centrifuge at above 4000 r/min at room temperature (20-25 °C) for 5min, until clear.
- 3 Take 50µL up-layer clear liquid, add 450 µL of the diluted redissolving solution, mix evenly for 30s.
- 4 Take 50 µL for further analysis.

Fold of dilution of sample: 20 Detection limit: 2ppb

Milk, milk powder

- 1 Take 2±0.05g sample, add 8mL 0.05M PB Buffer, shake for 5min, put it at 56°C water bath for 30min;
- 2 Centrifuge at above 4000 r/min at room temperature for 10 min.
- 3 Remove up-layer fat, take 50ul middle-layer clear liquid, add 450µL the diluted redissolving solution, mix properly for 30 seconds.
- 5 Take 50 µL for analysis.

Fold of dilution of sample:50 Detection limit: 5ppb

ELISA procedures

Instructions

1. Bring all reagents and micro-well strips to the room temperature (20-25°C).
2. Return all reagents to 2-8°C immediately after use.
3. The reproducibility of the ELISA analysis, to a large degree, depends on the consistency of plate washing.
plate washing. The correct operation of plate washing is the key point in the procedures of ELISA.
4. For the incubation at constant temperatures, all the samples and reagents must avoid light exposure, and each microplate should be sealed by the cover membrane.

Operation procedures

1. Bring test kit to the room temperature (20-25 °C) for at least 30 min, note that each reagent must be shaken evenly before use; put the required micro-well strips into plate frames. Re-sealed the unused microplate, stored at 2-8 °C, not frozen.
2. Solution preparation: dilute 40mL of the 20×concentrated washing buffer with deionized water at 1:19 (1 part 20× concentrated washing buffer + 19 parts deionized water), or prepare as quantity needed;
3. Numbering: number the micro-wells according to samples and standard solution; each sample and standard solution should be performed in duplicate; record their positions.
4. Add 50 μL of the sample and standard solution to separate duplicate wells, then add 50 μL of antibody working solution to each well, shake properly, seal the microplate with the cover membrane, and incubate at 25 °C for 30 min;
5. Pour liquid out of microwell, add 250 μL/well of washing buffer, then take out, repeat 5 times , each time for 30s, at last flap to dry with absorbent paper(if there are the bubbles after flapping, cut them with the clean tips);.
6. Add 100 μL of enzyme conjugate to each well, shake properly, seal the microplate with the cover membrane, and incubate at 25 °C for 30 min; continue as step 5 for washing.
7. Coloration: add 50 μL of the substrate A solution, then 50 μL of the B solution into each well. Mix gently by shaking the plate manually, and incubate at 25 °C for 15 min in the dark for coloration;
8. Determination: add 50 μL of the stop solution into each well. Mix gently by shaking the plate manually. Set the wavelength of the microplate reader at 450 nm to determine the OD value of every well. (Recommend to read the OD value at the dual-wavelength 450/630nm within 10 min).

Result judgment

There are two methods to judge the results; the first one is the rough judgment, while the second is the quantitative determination. Note that the OD value of the sample has a negative correlation with the content of Streptomycin

Qualitative determination

The concentration range (ng/mL) can be obtained from comparing the average OD value of the testing sample with that of the standard solution. Assuming that the OD value of the sample I is 0.3, and that of the sample II is 1.0, the OD value of standard solutions is: 2.243 for 0ppb, 1.816 for 0.1ppb, 1.415 for 0.3ppb, 0.74 for 0.9ppb, 0.313 for 2.7ppb, 0.155 for 8.1ppb, accordingly the concentration range of the sample I is 0.1 to 0.9ppb, and that of the sample II is 2.7 to 8.1ppb.

Quantitative determination

The mean values of the absorbance values obtained for the percentage of the average OD value (B) of the testing sample and the standard solution divided by the OD value (B₀) of the first standard solution (0 standard) and subsequently multiplied by 100%, that is,

$$\text{Percentage of absorbance value} = (B / B_0) \times 100\%$$

B—the average OD value of the sample or the standard solution

B₀—the average OD value of the 0 ng/mL standard solution

Draw the standard curve with the absorption percentages of the standard solutions and the semilogarithm values of the Streptomycin standard solutions (µg/L) as Y- and X-axis, respectively. Read the corresponding concentration of the testing sample from the standard curve by incorporating its absorption percentage into the standard curve. The resulting value is subsequently multiplied by the corresponding dilution fold, thus finally obtaining the Streptomycin concentration in the sample.

Using the professional analyzing software of this kit will be more convenient for the accurate and rapid analysis of a large amount of samples. (Please contact us for this software)

Precautions

1. The room temperature below 25 °C or the temperature of the reagents and the samples being not returned to the room temperature (20-25 °C) will lead to a lower standard OD value.
2. Dryness of the microplate in the washing process will be accompanied by the situations including the non-linear standard curves and the undesirable reproducibility. So continue to next step immediately after washing.
3. Mix evenly otherwise there will be the undesirable reproducibility.
4. The stop solution is the 2 M sulfuric acid solution, avoid contacting with the skin.
5. Do not use the kit exceeding its expiry date. The use of diluted or adulterated reagents from the kits will lead to the changes in the sensitivity and the detecting OD values. Do not exchange the reagents from the kits of different lot numbers to use.
6. Put the unused microplate into an auto-sealing bag to re-seal it. The standard substance and the colourless color former is light sensitive, and thus they cannot be directly exposed to the light.
7. Discard the colouration solution with any any color that indicates the degeneration of this solution. The detecting value of standard solution 1 (0 ppb) of less than 0.5 indicates its degeneration.
8. The optimum reaction temperature is 37 °C, and too high or too low temperatures will result in the changes in the detecting sensitivity and OD values.

Storage: store at 2-8 °C, not frozen.

Expiry date: 12 months; date of production is on box.

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