

## Chloramphenicol ELISA test Kit

Catalog No. LT70001AYSL

### 1. Principle

This test kit is based on the indirect competitive enzyme immunoassay for the detection of Chloramphenicol in the sample. The coupling antigen is pre-coated on the micro-well stripes. The Chloramphenicol in the sample and the coupling antigen pre-coated on the micro-well stripes compete for the anti-Chloramphenicol antibody. After the addition of the enzyme conjugate, the TMB substrate is added for coloration. The optical density (OD) value of the sample has a negative correlation with the Chloramphenicol in it. This value is compared to the standard curve and the Chloramphenicol concentration is subsequently obtained.

### 2. Technical specifications

**Sensitivity:** 15 ppt

**Incubation Temperature:** 25°C

**Incubation Time:** 30min~15min

**Detection limit:**

Tissue (method 1) .....15ppt

Tissue, egg (method 2).....30ppt

**Recovery rate**

Tissue, egg 95±25%

**Cross-reaction rate:**

Chloramphenicol..... 100%

Thiamphenicol..... < 0.1%

Florfenicol..... < 0.1%

### 3. Components

1	Micro-well strips	12 strips with 8 removable wells each	
2	7× standard solution (1mL each)	0ppt	15ppt 45ppt 135ppt 405ppt 1215ppt 10ppb
3	Enzyme conjugate	7ml	red cap
4	Antibody working solution	7ml	blue cap
5	Substrate A	7ml	white cap
6	Substrate B	7ml	black cap
7	Stop solution	7ml	yellow cap
8	20× concentrated washing buffer	40ml	white cap
9	2×concentrated redissolving solution	50ml	transparent cap
10	2-Nitrobenzaldehyde (C <sub>7</sub> H <sub>5</sub> NO <sub>3</sub> )	10ml	black cap

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CE Certified



ISO 9001



ISO 13485



ISO 14001



OHSAS 18001

#### 4. Materials required but not provided

##### Equipment:

- 1) microplate reader, homogenizer, nitrogen-drying device, vortex, centrifuge, measuring pipets, and balance (a sensibility reciprocal of 0.01 g), incubator.
- 2) **Micropipette:** single-channel 20-200  $\mu\text{L}$ , 100-1000  $\mu\text{L}$ , and multi-channel 30~300  $\mu\text{L}$ .
- 3) **Reagents:** Ethyl acetate, N-hexane, NaOH,  $\text{K}_2\text{HPO}_4 \cdot 3\text{H}_2\text{O}$ , HCl.

#### 5. Sample pre-treatment

##### **Instructions**

The following points must be dealt with before the pre-treatment of any kind of sample:

- 1) Only the disposable tips can be used for the experiments and the tips must be changed when used for absorbing different reagents;
- 2) Before the experiment, each experimental equipment must be checked to be clean and should be re-cleaned if necessary, in order to avoid the contamination which interferes with the experimental results.

##### **Solution preparation before sample pre-treatment**

- 1) Sample redissolving solution: The 2 $\times$ concentrated redissolving solution is diluted with deionized water at 1:1.
- 2) 0.1 M  $\text{K}_2\text{HPO}_4$  : dissolve 11.4g  $\text{K}_2\text{HPO}_4 \cdot 3\text{H}_2\text{O}$  in deionized water to 500mL.
- 3) 1 M HCl: dissolve 8.6mL HCl (approx 36.5%) in deionized water to 100mL.
- 4) 1 M NaOH: dissolve 4g NaOH in deionized water to 100mL.

##### **5.1 Tissue (Method 1)**

1. Take  $2 \pm 0.05$  g of the homogenized fresh sample into a 50ml centrifuge tube. Firstly add 3 mL deionized water, then add 6mL ethyl acetate, shake properly for 1 min, centrifuge at above 4000 r/min at room temperature (20-25 $^{\circ}\text{C}$ ) for 5 min.
2. Take 3mL of the up-layer liquid, blow to dry by nitrogen in 50-60 $^{\circ}\text{C}$ .
3. Dissolve the dry residues in 1 mL N-hexane, add 1 mL of the sample redissolving solution, mix for 30 seconds; centrifuge at above 4000 r/min at room temperature (20-25 $^{\circ}\text{C}$ ) for 5 min, remove the up-layer organic phase.
4. Take 50  $\mu\text{L}$  of the down-layer liquid for analysis.

##### **Fold of dilution of the sample: 1**

*(This method could not detect liver samples, and emulsification was easy to occur during treatment, which affected the detection results. (it was suggested to remove up-layer organic phase, then add 2ml N-hexane, mix it for 30s, centrifuge at above 4000 r/min at room temperature (20-25  $^{\circ}\text{C}$ ) for 10min; Remove the up-layer organic phase)*

## 5.2 Tissue, egg (Method 2)

- 1) Weigh  $1 \pm 0.05$ g of the homogenized sample, add 4mL of the distilled water, 0.5mL 1 M HCl and 100 $\mu$ L 2-Nitrobenzaldehyde ( $C_7H_5NO_3$ ) to each tube, shake properly for 2min;.
- 2) Incubate at **70 °C** by water bath for **20 minutes** (or incubate at 75 °C incubator for 25min).
- 3) Add 5mL 0.1M  $K_2HPO_4$ , 0.4mL 1M NaOH and 6mL ethyl acetate to each tube, shake for 30s.
- 4) Centrifuge at above 4000r/min at room temperature (20-25 °C ) for 5min ( if there is Emulsification or ethyl acetate layer is not enough for 3ml, incubate sample at 80 °C water bath for 10min and centrifuge repeatedly; or increase speed and extend time of centrifuge).
- 5) Transfer 3mL of the up-layer liquid into a new centrifugal tube and evaporate to dryness by nitrogen or air at 50 °C .
- 6) Dissolve the dry residues in 2mL N-hexane, add 1mL of the diluted redissolving solution, mix properly for 30 seconds, centrifuge at above 4000 r/min at room temperature (20-25 °C ) for 5 min; Remove up-layer N-hexane phase (if there is Emulsification, after removing up-layer N-hexane phase, incubate sample at 70 °C water bath for 10-20min, centrifuge repeatedly ).
- 7) Take 50  $\mu$ L of the down-layer for analysis.

**Fold of dilution of the sample: 2**

*(This method is all-in-one method, same sample preparation for AMOZ, AOZ, AHD, SEM and CAP etc.)*

## 6. ELISA procedures

### 6.1 Instructions

- 1 Bring all reagents and micro-well strips to the room temperature (20-25 °C) before use;
- 2 Return all reagents to 2-8 °C immediately after use;
- 3 The reproducibility of the ELISA analysis, to a large degree, depends on the consistency of plate washing. The correct operation of plate washing is the key point in ELISA the procedures;
- 4 For the incubation at constant temperatures, all the samples and reagents must avoid light exposure, and each microplate should be sealed by the cover membrane.

### 6.2 Operation procedures

1. Take out all the necessary reagents from the kit and place at the room temperature (20-25 °C) for at least 30 min. Note that each reagent must be shaken to mix evenly before use.
2. Take the required micro-well strips and plate frames. Re-sealed the unused microplate, store at 2-8 °C, not frozen.
3. Solution preparation: dilute 40 mL of the concentrated washing buffer (20 × concentrated) with the deionized water at 1:19 (1 part of 20X concentrated washing buffer + 19 parts of deionized water), or prepare as quantity needed.
4. Numbering: number the micro-wells according to samples and standard solution; each sample and standard solution should be performed in duplicate, record their positions.
5. Add 50  $\mu$ L of the sample or standard solution to separate duplicate wells; then add 50  $\mu$ L enzyme conjugate into each well, at last add 50  $\mu$ L of antibody working solution into each well. Mix gently by shaking the plate manually, seal the microplate with the cover membrane, and

**incubate at 25 °C for 30 min.**

6. Pour the liquid, wash the microplate with the diluted washing buffer at 250 µL/well for 4-5 times. Each time soak the well with the washing buffer for 15-30 sec, flap to dry with absorbent paper (if there are the bubbles after flapping, cut them with the clean tips).
7. Coloration: add 50 µL of the substrate A solution and then 50 µL of the B solution into each well. Mix gently by shaking the plate manually, and **incubate at 25 °C for 15 min at dark for coloration.**
8. Determination: add 50 µL of the stop solution into each well. Mix gently by shaking the plate manually. Set the wavelength of microplate reader at 450 nm to determine the OD value. (Recommend to read the OD value at the dual-wavelength 450/630 nm within 5 min).

## 7. Result judgment

There are two methods to judge the results; the first one is the rough judgment, while the second is the quantitative determination. Note that the OD value of the sample has a negative correlation with the content of Chloramphenicol.

### 7.1 Qualitative determination

The concentration range (ng/mL) can be obtained from the comparison the average OD value of the sample with that of the standard solution. Assuming that the OD value of the sample I is 0.3, and that of the sample II is 1.0, while those of the standard solutions are as the followings: 2.243 for 0ppt, 1.816 for 15ppt, 1.415 for 45ppt, 0.74 for 135ppt, 0.313 for 405ppt and 0.155 for 1215ppt, accordingly the concentration range of the sample I is 405 to 1215ppt, and that of the sample II is 45 to 135ppt.

### 7.2 Quantitative determination

The mean values of the absorbance values obtained for the average OD value (B) of the sample and the standard solution divided by the OD value (B<sub>0</sub>) of the first standard solution (0 standard) and subsequently multiplied by 100%, that is,

$$\text{Percentage of absorbance value} = \frac{B}{B_0} \times 100\%$$

B—the average OD value of the sample or the standard solution B<sub>0</sub>—  
the average OD value of the 0ng/mL standard solution

Draw the standard curve with the absorption percentages of the standard solution and the semilogarithm values of the Chloramphenicol standard solution (ng/mL) as Y- and X-axis, respectively. Read the corresponding concentration of the sample from the standard curve by incorporating its absorption percentage into the standard curve. The resulting value is subsequently multiplied by the corresponding dilution fold, thus finally obtaining the Chloramphenicol concentration in the sample.

Using the professional analyzing software of this kit will be more convenient for the accurate and

rapid analysis of a large amount of samples. (Please contact us for this software) .

### 8. Precautions

1. The room temperature below 25 °C or the temperature of the reagents and the samples being not returned to the room temperature (20-25 °C) will lead to a lower standard OD value.
2. Dryness of the microplate in the washing process will be accompanied by the situations including the non-linear standard curves and the undesirable reproducibility.
3. Mix every reagent and reaction mixture evenly and wash the microplate thoroughly, otherwise there will be the undesirable reproducibility.
4. The stop solution is the 2 M sulfuric acid solution, avoid contacting with the skin;
5. Put the unused microplate into an auto-sealing bag to re-seal it. The standard substance and the colourless color former is light sensitive, and thus they cannot be directly exposed to the light.
6. Do not use the kit exceeding its expiry date. The use of diluted or adulterated reagents from the kits will lead to the changes in the sensitivity and the detecting OD values. Do not exchange the reagents from the kits of different lot numbers to use.
7. Discard the colouration solution with any color that indicates the degeneration of this solution. The detecting value of the standard solution 1 (0 ppb) of less than 0.5 indicates its degeneration.
8. The optimum reaction temperature is 25 °C, and too high or too low temperatures will result in the changes in the detecting sensitivity and OD values.

### 9. Storage and expiry date

**Storage:** store at 2-8 °C, not frozen.

**Expiry date:** 12 months

**For Research Use Only.**

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