XL1Blue MAX

Catalog #	Package Size
1217-12	6x50 µl
1217-24	12x50 µl

Description

Intact Genomics (ig®) XL1 Blue Max electrocompetent E. coli cells offer the highest transformation efficiencies of ≥5 x 10^{10} cfu/µg plasmid DNA which are ideal for applications requiring high transformation efficiencies, such as with cDNA or gDNA library construction. These cells have the capability to allow for the preparation of high quality plasmid DNA, single strand rescue of phagemid DNA and Blue/white screening. XL1 Blue Max cells provide superb transformation efficiency, significantly higher than any competitors similar product, allowing for increased opportunity for experimental success.

Specifications

Competent cell type: ElectroCompetent
Species: *E. coli*
Format: Tubes
Transformation efficiency: ≥ 2.5 x 10^{10} cfu/µg pUC19 DNA
Blue/white screening: Yes
Shipping condition: Dry ice

Reagents Needed for One Reaction

ig® XL1 Blue Max electrocompetent cells: 25 µl
DNA (or pUC19 Control, 10 pg/µl): 1 µl
Recovery medium: 1 ml

Storage

ig® XL1 Blue Max electrocompetent cells: -80 ºC
pUC19 control DNA: -20 ºC
Recovery medium: 4 ºC

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Product Benefits

ig® XL1 Blue Max electrocompetent cells have the following features:

- XL1 Blue Max cells are tetracycline resistant.
- XL1 Blue Max cells are endonuclease (endA) deficient, which greatly improves the quality of miniprep DNA.
- XL1 Blue MAx cells recombination (recA) deficient, improving insert stability.
- Cleavage of cloned DNA by the EcoK endonuclease system is prevented by the hsdR mutation.
- Bue-white color screening via the laclq ZΔM15 gene on the F´ episome.

Genotype

recA1 endA1 gyrA96 thi-1 hsdR17 supE44 relA1 lac [F´ proAB laclq ZΔM15 Tn10 (Tetr)].

Quality Control

Transformation efficiency is tested by using the pUC19 control DNA supplied with the kit and the high efficiency transformation protocol listed below. Transformation efficiency should be ≥5 x 10^{10} CFU/µg pUC19 DNA.

Untransformed cells are tested for appropriate antibiotic sensitivity.

General Guidelines

Follow these guidelines when using ig® XL1 Blue Max ElectroCompetent *Cells:*

- Handle competent cells gently as they are highly sensitive to changes in temperature or mechanical lysis caused by pipetting.
- Thaw competent cells on ice, and transform cells immediately following thawing. After adding DNA, mix by tapping the tube gently. Do not mix cells by pipetting or vortexing.

Note: A high-voltage electroporation apparatus such as Bio-Rad Gene Pulser II #165-2105, capable of generating

Transformation Protocol

Use this procedure to transform ig® XL1 Blue Max electrocompetent cells. Do not use these cells for chemically transformation.

1) Place sterile cuvettes and microcentrifuge tubes on ice.
2) Remove competent cells from the -80 °C freezer and thaw completely on wet ice (10-15 minutes).
3) Aliquot 1 µl (1 pg-10 ng) of DNA to the chilled microcentrifuge tubes on ice.
4) When the cells are thawed, add 25 µl of cells to each DNA tube on ice and mix gently by tapping 4-5 times. For the pUC19 control, add 1 µl of (10 pg/µl) DNA to the 25 µl of cells on ice. Mix well by tapping. Do not pipette up and down or vortex to mix, this can harm cells and decrease transformation efficiency.
5) Pipette 26 µl of the cell/DNA mixture into a chilled electroporation cuvette without introducing bubbles. Quickly flick the cuvette downward with your wrist to deposit the cells across the bottom of the well and then electroporate.
6) Immediately add 974 µl of Recovery Medium or any other medium of choice to the cuvette, pipette up and down three times to re-suspend the cells. Transfer the cells and Recovery Medium to a culture tube.
7) Incubate tubes at 37 °C for 1 hour at 210 rpm.
8) Dilute the cells as appropriate then spread 20-200 µl cells onto a pre-warmed selective plate. For the pUC19 control, plate 50 µl of diluted transformants onto an LB plate containing 100 µg/ml ampicillin. Use sterilized spreader or autoclaved ColiRoller™ plating beads to spread evenly.
9) Incubate the plates overnight at 37 °C.

Example Calculation of TE

-Example not strain specific-

Transformation Efficiency (TE) is defined as the number of colony forming units (cfu) produced by transforming 1µg of plasmid into a given volume of competent cells.

TE = Colonies/µg/Dilution

Transform 1 µl of (10 pg/µl) pUC19 control plasmid into 25 µl of cells, add 950 µl of Recovery Medium. Dilute 10 µl of this in 990 µl of Recovery Medium and plate 50 µl. Count the colonies on the plate the next day. If you count 100 colonies, the TE is calculated as follows:

Colonies = 100
µg of DNA = 0.00001
Dilution = 50/1000 x 10/1000 = 0.0005
TE = 100/.00001/.0005 = 2.0×10^{10}

Related Products

- ig® 5-Alpha Chemically Comp. Cells (Cat.# 1031-12)
- ig® 10B Chemically Comp. Cells (Cat.# 1011-12)
- T4 DNA Ligase (Cat.# 3212)
- i7® High Fidelity DNA Polymerase (Cat.# 3254)
- igFusion™ Cloning Kit (Cat.# 4111)

Technical Support

Intact Genomics is committed to supporting the worldwide scientific research community by supplying the highest quality reagents. Each new lot of our products is tested to ensure they meet the quality standards and specifications designated for the product.
Please follow the instructions carefully and contact us if additional assistance is needed. We appreciate your business and your feedback regarding the performance of our products in your applications.

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